

2 α -Methylhopanoids: First Recognition in the Bacterium *Methylobacterium organophilum* and Obtention via Sulphur Induced Isomerization of 2 β -Methylhopanoids.

An account for their presence in sediments.

P Stampf, D Herrmann, P Bisseret and M Rohmer*

Ecole Nationale Supérieure de Chimie de Mulhouse, 3 rue A Werner, 68093 Mulhouse Cedex, France

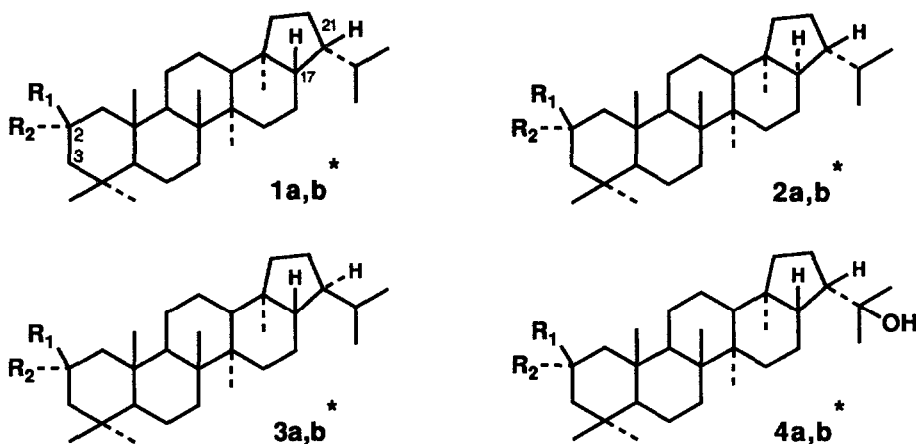
(Received in Belgium 17 May 1991)

Key Words Hopanoids, methylhopanoids, 2 α -methylplopterol, sulphur-induced isomerization, bacterial triterpenoids

Abstract First recognition of 2 α -methylplopterol from the bacterium *Methylobacterium organophilum* and sulphur induced isomerization of 2 β -methylhopanoids into their 2 α -methyl isomers suggest a dual origin for the sedimentary 2 α -methylhopanoids

Triterpenoids from the hopane family isolated from living organisms ("biohopanoids") are typically derived from the C₃₀ 17 β ,21 β framework 1 (Scheme 1) ¹ They are the precursors of the many hopanoids encountered in sediments ("geohopanoids") which often possess the thermodynamically more stable skeleton 2, and to a lesser extent 3 of respective 17 α ,21 β and 17 β ,21 α configurations ² Beside these major compounds, minor members possessing an additional methyl group attached at position 2 β or 3 β of their 17 β ,21 β skeleton have been isolated from a few bacteria scattered through several taxonomic groups ^{1a,3} These are thus the putative precursors of the widespread geohopanoids characterized by an additional methyl group at position 2 or 3 of their 17 α ,21 β skeleton. Only recently have the configurations at C-2 and C-3 been discussed. In the case of 3-methylhopanoids, compounds bearing the thermodynamically favoured equatorial 3 β -methyl group were only encountered, whereas a mixture of both 2 α - and 2 β -methylhopanoids appeared in younger sediments ^{4,5} In more mature geological samples, only the 2 α -methyl isomers were found, ^{4,5} illustrating thus the transformation into the most stable series during the diagenesis as early recognized for the apparent 17 β /17 α epimerization ² To account for this latter isomerization, we pointed out recently the good epimerization power of sulphur, an element often present in sediments, at high temperatures ⁶ We present here further heating simulations in liquid sulphur to investigate the 2 β -methyl/2 α -methyl isomerization using mainly 2 β -methyl-17 α -hopane 2a, with the

thermodynamically most stable configuration at C-17, as a model compound and the first identification of 2 α -methyl diplopterol from the bacterium *Methylobacterium organophilum*.



Scheme 1. *Non indexed figures refer to $R_1=R_2=H$, a indexed ones to $R_1=CH_3$, $R_2=H$ and b indexed ones to $R_1=H$, $R_2=CH_3$

2 α -METHYLDIPLOPTEROL IN *Methylobacterium organophilum* IDENTIFICATION AND BIOSYNTHESIS

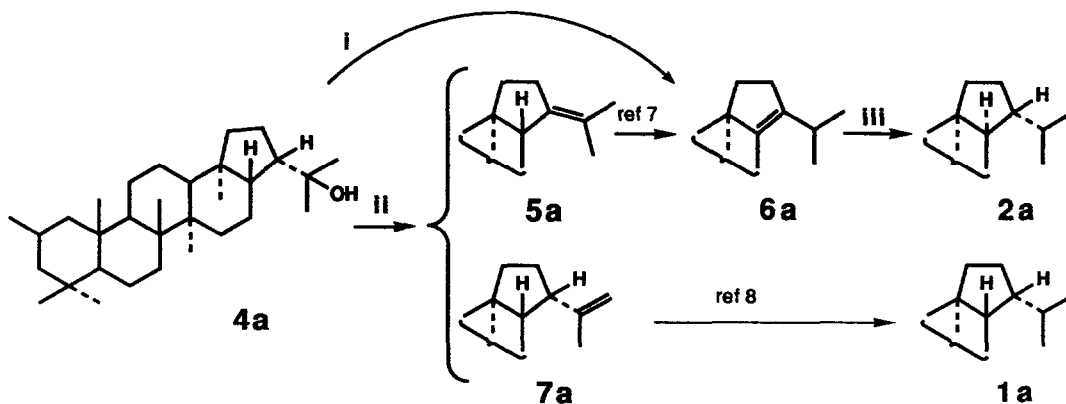
With larger amounts (20mg) of tlc-purified 2 β -methyl diplopterol **4a**, isolated from *M. organophilum* and reported as pure when the bacterium was grown on the Hestrin and Schramm medium,^{3a} we repeated the ¹³C-nmr analysis and detected ca 5% of its 2 α -methyl isomer **4b** by comparison with the spectrum now available of synthetic 2 α -methyl diplopterol. Further improvement of the gc analyses permitted also to detect this minor compound possessing a slightly longer retention time than that of 2 β -methyl diplopterol.

Methylation of hydrocarbon skeletons occurs usually via an olefinic precursor, S-adenosylmethionine being the methyl donor, as shown for instance for the methylation reactions of phytosterol side-chains. Feeding experiments with (2H₃)-methylmethionine showed that the labelled methyl group was transferred with its three deuterium atoms into 2 β -methyl diplopterol.^{3c} Although an unsaturated hopanoid could not be detected until now, this confirmed the role of methionine as precursor of the methyl donor and excluded intermediates with methylene or cyclopropyl groups in this biosynthetic pathway. Reinvestigation by gc-ms of this deuterated 2 β -methyl diplopterol sample using the new gc procedure permitted again to identify 2 α -methyl diplopterol. Next to non-labelled 2 α -methyl diplopterol synthesized from non-labelled carbon source, only the trideuterated analogue arising from labelled methionine, and no mono- and dideuterated compounds, could be detected. The conclusions concern-

ning the structures of possible intermediates for the biosynthesis of 2 α -methylhopanoids are thus similar to those described for the formation of their 2 β -methyl isomers. It has still to be determined whether there are two similar, but separate and stereoselective biosynthetic pathways, or one single enzymatic reaction sequence with low stereoselectivity.

SYNTHESIS OF 2 β -METHYL AND 2 α -METHYLHOPANOIDS

2 β -Methyl-17 β - and 17 α -hopanes **1a** and **2a**, both accompanied by 5% of their 2 α -methyl isomer, *i.e.* reflecting the natural isomeric ratio at C-2 in *M. organophilum*, were obtained from natural 2 β -methyl diplopterol **4a** according to classical methods (Scheme 2)



Scheme 2 Synthesis of 2 β -methyl-17 β - and 17 α -hopanes **1a** and **2a** starting from 2 β -methyl diplopterol **4a** isolated from *M. organophilum*, each compound is accompanied by traces (5%) of 2 α -methyl isomer **1**, HClO₄, HCO₂H, CHCl₃, 20°C, 10min, ii, SOCl₂, Py, 15 min, 0°C, iii, H₂, Pd(OH)₂, AcOH, AcOEt, HClO₄, 18h, 20°C

After thionyl chloride dehydration of 2 β -methyl diplopterol **4a** in the presence of pyridine,⁹ silver nitrate tlc purification yielded the two olefins **5a** and **7a** in a 2:3 proportion, giving thus comparable results as the Ac₂O/K₂CO₃ mediated dehydration reported on diplopterol **4**.¹⁰ 2 β -Methyl diploptene **7a** on the one hand was directly hydrogenated using Adam's catalyst into the required 2 β -methyl-17 β -hopane **1a**.⁸ 2 β -Methylhop-21-ene **5a** on the other hand was converted to the $\Delta^{17(21)}$ -unsaturated compound **6a** under acidic conditions,⁷ which then was hydrogenated to the 17 α isomer **2a** using either Adam's or Pearlman's catalyst, both under strong acidic conditions.¹¹ Use of a palladium catalyst proved fruitful, as recognized by Summons and Jahnke with Pd/C,^{4a} yielding a better stereospecific conversion into **2a**. When required, **4a** was efficiently directly dehydrated into **6a** in the presence of perchloric acid.

2 α -Methyl-17 β - and 17 α -hopanes **1b** and **2b** were prepared from 22-hydroxyhopan-3-one **8a** according to the improved methylation procedure shown in Scheme 3. The initial method on small quantities of **8a** (<0.1mmole)^{3a} yielding rather irrepro-

Table 1: Heating of hopanoids 1a and 2a in liquid sulphur ^a

Entry	Compound	Hopanoid /Sulphur (mg)	T(°C)	t(h)	Yield ^b %	Composition ^c %
1	2a	3 / 30	200	3 h	65	2a (90%), 2b (5%), unknowns (5%)
2	2a	3 / 30	240	3 h	50	2a (80%), 2b (15%), unknowns (5%)
3	2a	3 / 30	240	24h	35	2a (80%), 2b (15%), unknowns (5%)
4	1a	6 / 30	240	3 h	45	1a,b (6%) ^d , 2a,b (75%) ^d , 3a,b (17%) ^d , minor compounds (2%) ^e

a, for experimental conditions, see ref 6, b, for apolar compounds, the rest represents a more polar brown material, c, analyses performed thanks to synthetic reference materials, % estimated both by gc and ¹³C-nmr for entries 1-3 and by gc only for entry 4, d, 2 α -methyl/2 β -methyl isomeric ratio = ca 15 85 (gc), e, see ref 6

DISCUSSION

The detection of 2 α -methylhopanoids in *M. organophilum* represents the first record in a living organism of a series so far reported only in some sediments. 2 α -Methylhopanoids have only been so far reported from hypersaline and carbonate sediments⁴ and from bitumen samples from archeological sites⁵. This apparent lack of methylation stereospecificity has to be compared to the usual occurrence of both (24*R*)- and (24*S*)-methylsterols in many plant species. Indeed, the biosynthesis of 2 β -methylhopanoids was shown to involve *L*-methionine,^{3c} most probably *via* *S*-adenosylmethionine as a methyl donor, like in higher plants. As it is reasonable to anticipate the presence of 2 α -methylhopanoids in other prokaryotes, this finding in itself could account for the occurrence of this series in sediments but not for its predominance compared to 2 β -methylhopanoids, unless the formers appear much more widespread in bacteria. To investigate the possibility of diagenetic-induced epimerization of 2 β -methyl into the 2 α -methyl group, we extended our experiments of sulphur-induced isomerization at C-17 to carbon C-2 of methylhopanoids. The epimerization power of sulphur in this case proved to be modest (ca.10%) in response to probably less clear-cut kinetic and thermodynamic criteria. Indeed the attacked 2 α C-H bond is not as well exposed as the 17 β C-H bond, and the sterical decompression resulting from the 2 β -methyl/2 α -methyl conversion is probably not so important, as the ring-A may adopt a twisted conformation to minimize the steric interactions of its 2 β -methyl group with the neighbouring 4 β - and 10 β -methyl groups.¹⁵ The mere existence of this isomerization could even be questioned as the enhancement in 2 α -methyl isomer after the heating experiments could only reflect a greater tendency of the 2 β -methyl isomer to decompose to more polar compounds (see Table 1). The fact, however, that the 2 α -methyl/2 β -methyl isomeric ratio does not increase on prolonged heating at 240°C (Table 1, entry 3) precludes this possibility. In the case of hopanoid 1a, the slowness of the 2 β -methyl/2 α -methyl epimerization compared to the 17 β ,21 β /17 α ,21 β and to a lesser extent 17 β ,21 β /17 β ,21 α ones is particularly enlightened as hopanoid composition after the heating experiment essentially reflects the epi-

merization at C-17. Two features have to be kept in mind. The reaction conditions we used (molten sulphur) are less likely to occur in sediments and the apparent isomerization we observed might be the result of a more complex process involving the formation of functionalized intermediates and their decomposition into the detected end products.¹⁶

Our results suggest a dual origin for the presence of 2 α -methylhopanoids in sediments: *i.e.* the direct incorporation of prokaryotic 2 α -methylhopanoids and/or abiotic isomerization of 2 β -methyl isomers. However, several questions remain, considering the reported predominance of sedimentary 2 α -methyl isomers on the one hand, and the low epimerization power of sulphur on the other hand. Do 2 α -methylhopanoids have a broad distribution in prokaryotes? Does diagenesis involve other non sulphur-induced epimerization processes? Could it be only that the presence of 2 β -methyl and 2 α -methylhopanoids in sediments has been much overlooked? Indeed, in sedimentary mixtures of methylated and non-methylated hopanoids, detection of 2 β -methylhopanoids, especially in small amounts, appears delicate as, contrary to their 2 α and 3 β isomers, they possess gc retention times comparable to those of non-methylated hopanoids.

ACKNOWLEDGEMENTS

We thank Mrs E Krempp and Dr D. Le Nouen for nmr measurements as well as Mr P. Wehrung for ms measurements. This work was supported by the Centre National de la Recherche Scientifique (Unité Associée 135) and by the Ministère de l'Education Nationale (Prolongement International de la Recherche).

EXPERIMENTAL

General

Most of the analytical procedures and separation schemes were as described previously.^{1a} Gc analyses were performed on a CARLO ERBA Fractovap 4160 chromatograph equipped with a flame ionization detector fitted with a DB-1 surface-bonded phase fused silica capillary column (30m, film thickness=0.1 μ m) using a detector temperature of 310°C and a standard temperature program from 50°C to 220°C (20°C/min) and from 220°C to 310°C (4°C/min) with a hydrogen pressure of 0.7 kg cm⁻². When required (Table 1), nearly base line separation between 2 α -methyl and 2 β -methylhopanes isomers was obtained using a slower temperature program *i.e.* from 50°C to 175°C at 20°C/min and from 175°C to 310°C at 0.5°C/min. Electronic impact gc/ms analyses were performed at 70eV on a LKB 9000S spectrometer as previously reported¹⁷ or on a KRATOS MS80RF mass spectrometer with a source temperature of 280°C and a DB-5 (30m x 0.25mm) capillary column. ¹H-nmr and ¹³C-nmr spectra were recorded in C²HCl₃ on BRUKER W200 or W400 or ACF250 apparatuses using CHCl₃ (δ =7.260ppm) as internal standard for ¹H-nmr or ¹³C²HCl₃ (δ =77.03ppm) for ¹³C-nmr. Melting points were measured after recrystallization from CH₂Cl₂/CH₃OH using a REICHERT-JUNG micro hot-stage and are uncorrected.

3,22-Bistrimethylsiloxy-hop-2-ene 9

To a solution of hydroxyhopanone **8a** (105mg) in DMF (2ml) and triethylamine (1ml), chlorotrimethylsilane (1ml) was added. The resulting mixture from which a pale yellow solid (presumably triethylamine hydrochloride) separated immediately was heated for 15h at 50°C. After cooling, dilution with ether (10ml) and washing with aq NaHCO₃ (10ml), the organic layer was dried (Na₂SO₄) and concentrated, yielding the required bistrimethylsilyl ether **9** (125mg) next to 22-trimethylsiloxy-hopan-3-one **8b** (10mg) after flash chromatography (Si-gel, 5% EtOAc-cyclohexane)

9, ¹H-nmr (200MHz), δ 0.0094 (9H, s), 0.179 (9H, s), 0.737 (3H, s), 0.848 (3H, s), 0.940 (3H, s), 0.971 (3H, s), 1.000 (3H, s), 1.146 (3H, s), 1.245 (3H, s), 1.962 (1H, dd, J=16 & 6.6Hz), 2.000 (2H, m), 4.573 (1H, dd, J=1.8 & 6.6Hz) ppm

8b, m p =194-195°C, ¹H-nmr (200MHz); δ 0.093 (9H, s), 0.735 (3H, s), 0.919 (3H, s), 0.942 (3H, s), 0.990 (3H, s), 1.020 (3H, s), 1.073 (3H, s), 1.144 (3H, s), 1.245 (3H, s), 2.382 (1H, ddd, J=4.5 & 7.5 & 15.5Hz), 2.507 (1H, ddd, J=7 & 9.5 & 15.5Hz) ppm.

2 α - and 2 β -Methylhopan-22-ol-3-one 10b and 10a

Benzyltrimethylammonium fluoride (105mg) was dried overnight in THF (2ml) in the presence of 4Å molecular sieve (600mg) under an argon atmosphere. To this suspension, a solution of bistrimethylsilyl ether **9** (50mg) in THF (2ml) and CH₃I (30 μ l) were added. After stirring of the mixture for 1h at room temperature and 4h at 50°C, filtration and removal of the solvent, tlc purification (Si-gel, 5% EtOAc-toluene), next to unreacted starting compound **9** (R_f=0.82, 4mg) and trimethylsilyl ether **8b** (R_f=0.41, 15mg, 34%), yielded 22-O-trimethylsilylether of 2-methyl-22-hydroxyhopan-3-one (R_f=0.53, 21mg). A solution of the latter (18mg) in MeOH (1ml) and THF (1ml) was stirred, after addition of HCl 6N (15 μ l), for 2h at room temperature. Removal of the solvent and tlc purification (Si-gel, 5% EtOAc-toluene), besides unreacted starting product (1.4mg), afforded 2-methylhydroxyhopanone **10a** and **10b** (15 mg) which appeared to be a 4:1 mixture of 2 α :2 β methyl isomers (¹³C-nmr). The 2 α - and 2 β -methylhydroxyhopanone **10b** and **10a** were obtained pure after C₁₈ reverse phase hplc purification using MeOH:H₂O, 92:8, v/v as eluent.

10a, ¹H-nmr (250MHz), δ 0.655 (3H, s), 0.776 (3H, s), 0.944 (3H, s), 0.975 (3H, d, J=6.4Hz), 0.988 (3H, s), 1.023 (3H, s), 1.056 (3H, s), 1.180 (3H, s), 1.210 (3H, s), 2.232 (1H, m), 2.804 (1H, ddq, J=6.4 & 9.4 & 11.0Hz) ppm

10b, ¹H-nmr (250MHz), δ 0.759 (3H, s), 0.929 (3H, s), 1.016 (3H, d, J=6.4Hz), 1.018 (3H, s), 1.036 (3H, s), 1.062 (3H, s), 1.106 (3H, s), 1.180 (3H, s), 1.211 (3H, s), 2.229 (1H, m), 2.752 (1H, quintuplet, J=6.4Hz) ppm

2 α -Methyldiplopterol 4b

2 α -Methylhydroxyhopanone **10b** was converted to 2 α -methyldiplopterol **4b** after Wolff-Kishner reduction as already described¹²

4b m p =194-196°C, ¹H-nmr (200MHz) see ref.3a, ¹³C-nmr (63MHz), δ 16.14 (C-28), 16.65 (C-25), 16.74 (C-27), 17.05 (C-26), 18.61 (C-6), 20.91 (C-11), 21.95 (C-16), 22.22 (C-24), 23.15 (C-31), 23.90 (C-2), 24.11 (C-12), 26.62 (C-20), 28.72 (C-29), 30.86 (C-30), 33.25 (C-7), 33.47 (C-23), 33.94 (C-4), 34.38 (C-15), 38.05 (C-10), 41.22 (C-19), 41.84 (C-14), 41.90 (C-8), 44.09 (C-18), 49.57 (C-3), 49.81 (C-13), 50.33 (C-9), 51.11 (C-21), 51.24 (C-1), 53.90 (C-17), 55.74 (C-5), 73.99 (C-22) ppm

2 β -Methyl-17 β -hop-21-ene 5a and 2 β -methyl-17 β -hop-22(29)-ene 7a

To a solution of tertiary alcohol **4a** (20mg) in a 2:1 mixture of CH₂Cl₂/pyridine (1.5ml) was added at 0°C under stirring freshly distilled thionyl chloride (100 μ l). After 15min at 0°C, the medium was quenched over aq. Na₂CO₃ and diluted with pet ether. The organic layer was dried (Na₂SO₄), evaporated and purified by tlc (Si-gel, 10% AgNO₃, pet ether-toluene, 95:5) to give **5a** (R_f=0.45, 6mg), **7a** (R_f=0.25, 8mg) next to traces of **6a** (R_f=0.35, 0.3mg)

5a, m.p.=173-174°C, ¹H-nmr (400MHz), δ 0.599 (3H, d, J=0.7Hz), 0.840 (3H, d, J=6.3Hz), 0.845 (3H, s), 0.900 (3H, s), 0.909 (3H, s), 0.968 (6H, s), 1.589 (3H, s), 1.741 (3H, s), 2.2 (3H, s) ppm

ms m/z 424 (M⁺, 78%), 409 (18%), 381 (52%), 355 (64%), 313 (8%), 245 (22%), 219 (10%), 217 (11%), 205 (82%), 189 (100%), 177 (18%), 175 (22%), 161 (51%).

7a, m.p.=202-203°C; ¹H-nmr (400MHz); δ 0.732 (3H, s), 0.833 (3H, d, J=7Hz), 0.838 (3H, s), 0.891 (3H, s), 0.900 (3H, s), 0.943 (3H, s), 0.957 (3H, s), 1.758 (3H, s), 2.69 (1H, dd, J=7 & 9Hz), 4.788 (2H, s) ppm

ms m/z 424 (M⁺, 32%), 409 (10%), 315 (6%), 313 (8%), 218 (10%), 205 (80%), 189 (100%), 177 (8%), 175 (7%), 163 (7%), 161 (16%)

2 α -Methyl-17 β -hop-21-ene 5b and 2 α -methyl-17 β -hop-22(29)-ene 7b

Preparation of the two methylhopenes **5b** and **7b** was realized as described for **5a** and **7a** (see above)

5b, m.p.=169-170°C, ¹H-nmr (400MHz); δ 0.583 (3H, d, J=0.8Hz), 0.800 (3H, s), 0.827 (3H, s), 0.829 (3H, d, J=6.2Hz), 0.852 (3H, s), 0.959 (3H, s), 0.965 (3H, s), 1.577 (3H, s), 1.730 (3H, s) ppm

ms m/z 424 (M⁺, 35%), 409 (8%), 381 (30%), 355 (36%), 245 (27%), 205 (100%), 189 (83%), 161 (62%).

7b, m.p.=205-206°C; ¹H-nmr (400MHz); δ 0.716 (3H, d, J=1Hz), 0.792 (3H, s), 0.818 (3H, s), 0.823 (3H, d, J=6.2Hz), 0.843 (3H, s), 0.932 (3H, s), 0.955 (3H, s), 1.748 (3H, s), 2.677 (1H, dt, J=7 & 8Hz), 4.77 (2H, s) ppm

ms m/z 424 (M⁺, 14%), 409 (6%), 381 (4%), 205 (100%), 189 (99%), 177 (7%), 175 (9%), 163 (10%), 161 (20%)

2 β -Methylhop-17(21)-ene 6a from 4a

After dissolution of 2 β -methylhop-17(21)-ene **4a** (15mg) in a minimum amount of CHCl₃ (ca 0.3ml), HCO₂H (0.3ml) and 70% HClO₄ (50 μ l) were added under stirring. The biphasic medium was stirred at room temperature vigorously for 10min, quenched over aq. NaHCO₃ and diluted with pet ether. After drying (Na₂SO₄), evaporation of the solvents and tlc purification (Si-gel, 10% AgNO₃, 5% toluene-pet ether), **6a** (R_f=0.35, 13.5mg) was obtained as a colourless solid

6a, m.p.=138-139°C, ¹H-nmr (400MHz), δ 0.838 (3H, s), 0.84 (3H, d, J=6.5Hz), 0.847 (3H, s), 0.898 (3H, s), 0.916 (3H, s), 0.922 (3H, s), 0.925 (3H, d, J=6.7Hz), 0.984 (3H, d, J=6.8Hz), 1.041 (3H, s), 1.92 (1H, m), 2.11 (1H, dd, J=9 & 15Hz), 2.26 (1H, dt, J=6.5 & 15Hz), 2.65 (1H, sept, J=7Hz) ppm

ms m/z 424 (M⁺, 57%), 409 (18%), 381 (100%), 245 (42%), 205 (37%), 203 (12%), 189 (24%), 175 (24%), 161 (38%)

2 α -Methylhop-17(21)-ene 6b

Methylhopene **6b** was prepared from **5b** as previously reported on its non-methylated homologue ⁷

6b, m p = 147-149°C; ¹H-nmr (400MHz), δ 0.795 (3H, d, J=6Hz), 0.833 (6H, s and 3H, d, J=6Hz), 0.851 (3H, s), 0.914 (3H, d, J=6.9Hz), 0.923 (3H, s), 0.974 (3H, d, J=6.9Hz), 1.034 (3H, s), 2.2 (2H, m), 2.64 (1H, sept, J=6.9Hz) ppm.

ms m/z 424 (M⁺, 22%), 409 (13%), 381 (79%), 245 (64%), 205 (65%), 189 (34%), 180 (45%), 161 (75%), 135 (100%)

2 α -and 2 β -Methyl-17 β -hopanes 1b and 1a

Both methylhopanes **1a** and **1b** were prepared by hydrogenation of **7a** and **7b** as previously described ⁸

1a, m p = 195-196°C; ¹H-nmr (400MHz), δ 0.702 (3H, s), 0.805 (3H, d, J=6.2Hz), 0.824 (3H, d, J=6.4Hz), 0.826 (3H, s), 0.881 (3H, d, J=6.2Hz), 0.824 (3H, d, J=6.4Hz), 0.826 (3H, s), 0.881 (3H, s), 0.889 (3H, s), 0.926 (3H, d, J=7Hz), 0.935 (6H, s) ppm

ms m/z 426 (M⁺, 3%), 411 (4%), 383 (6%), 205 (42%), 191 (100%), 163 (12%)

1b, m p = 213-214°C, ¹H-nmr (400MHz), δ 0.694 (3H, s), 0.791 (3H, s), 0.799 (3H, d, J=6.5Hz), 0.815 (3H, s), 0.822 (3H, d, J=6.5Hz), 0.844 (3H, s), 0.924 (3H, d, J=6.5Hz), 0.934 (3H, s), 0.943 (3H, s) ppm

ms. m/z 426 (M⁺, 4%), 411 (4%), 383 (6%), 205 (48%), 191 (100%), 163 (10%)

2 β -Methyl-17 α ,21 β -hopane 2a

After dissolution of the tetrasubstituted olefin **6a** (10mg) in a 1:1 mixture of AcOEt:AcOH (6ml), 70% HClO₄ was added (0.15ml) followed by palladium hydroxide (30mg). After 20h of stirring at 20°C under hydrogen (1atm), the mixture was quenched over aq. NaHCO₃, diluted with pet ether, dried (Na₂SO₄) and evaporated to dryness giving **2a** (R_f=0.85, 6mg) as a colourless solid after tlc purification (Si-gel, 10% AgNO₃; pentane)

2a, m p = 138-139°C, ¹H-nmr (250MHz), δ 0.815 (3H, d, J=6.5Hz), 0.824 (3H, s), 0.832 (3H, d, J=6.5Hz), 0.840 (3H, s), 0.892 (3H, s), 0.895 (3H, d, J=6.5Hz), 0.907 (3H, s), 0.925 (3H, s), 0.946 (3H, s), 0.995 (3H, s) ppm.

ms m/z 426 (M⁺, 8%), 411 (9%), 383 (2%), 231 (4%), 218 (11%), 205 (100%), 191 (48%), 177 (8%), 163 (22%).

2 α -Methyl-17 α ,21 β -hopane 2b

Methylhopane **2b** was prepared from **5b** as already described for the non-methylated series ^{7,11}

2b, m p = 88-89°C, ¹H-nmr (400MHz); δ , 0.798 (3H, s), 0.806 (3H, d, J=6.5Hz), 0.829 (3H, s), 0.829 (3H, d, J=6Hz), 0.848 (3H, s), 0.893 (3H, d, J=6.5Hz), 0.956 (3H, s), 0.992 (3H, s) ppm

ms m/z 426 (M⁺, 8%), 411 (9%), 383 (2%), 231 (4%), 218 (11%), 205 (100%), 191 (48%), 177 (8%), 163 (22%)

2 α -and 2 β -Methyldiplopterols 4b and 4a from *Methylobacterium organophilum*

2 α -Methyldiplopterol 4b was identified in trace amount (5%) in the ¹³C-nmr spectrum of a sample of 2 β -methyldiplopterol (20mg) previously isolated from *M organophilum*^{3a} by comparison with the spectrum of synthetic 2 α -methyldiplopterol of reference

2-Methyldiplopterols from *M. organophilum*

¹³C-nmr (63MHz), δ 16.14 (4b, C-28), 16.28 (4a, C-27 and C-28), 16.64 (4b, C-25), 16.74 (4b, C-27), 16.94 (4a, C-26), 17.04 (4b, C-26), 18.61 (4b, C-6), 19.91 (4a, C-6), 20.89 (4b, C-11), 21.75 (4a, C-25), 21.86 (4a, C-11), 21.94 (4a and 4b, C-16), 22.21 (4b, C-24), 23.15 (4b, C-31), 23.21 (4a, C-31), 23.89 (4b, C-2), 24.10 (4b, C-12), 24.42 (4a, C-20), 26.61 (4b, C-20), 28.70 (4a and 4b, C-29), 30.84 (4a and 4b, C-30), 31.03 (4a, C-23), 32.44 (4a, C-4), 32.47 (4a, C-7), 33.24 (4b, C-7), 33.46 (4b, C-23), 33.93 (4b, C-4), 34.36 (4b, C-15), 34.40 (4a, C-15), 37.78 (4a, C-10), 38.04 (4b, C-10), 41.21 (4b, C-19), 41.29 (4a, C-19), 41.82 (4b, C-14), 41.88 (4a, C-14 and 4b, C-8), 42.00 (4a, C-8), 44.08 (4b, C-18), 44.10 (4a, C-18), 45.15 (4a, C-1), 49.56 (4a and 4b, C-13), 49.70 (4a, C-3), 49.80 (4b, C-13), 50.27 (4a, C-9), 50.31 (4b, C-9), 51.10 (4a and 4b, C-21), 51.23 (4b, C-1), 53.88 (4b, C-17), 53.92 (4a, C-17), 55.72 (4b, C-5), 73.99 (4a and 4b, C-22) ppm

REFERENCES

- 1 (a) M Rohmer, P Bouvier-Navé, and G Ourisson, *J Gen Microbiol*, **1984**, *130*, 1137, (b) G Ourisson, M Rohmer, and K Poralla, *Ann Rev Microbiol*, **1987**, *41*, 301
- 2 (a) G Ourisson, P Albrecht, and M Rohmer, *Pure Appl Chem*, **1979**, *51*, 709, (b) M Rohmer, M Dastillung, and G Ourisson, *Naturwissenschaften*, **1980**, *67*, 456, (c) A Ensminger, A Van Dorsselaer, C Spycykerelle, P Albrecht, and G Ourisson, in *Advances in Organic Geochemistry* (eds B Tissot, and F Biener) Technip, Paris, **1973**, pp 245-260, (d) W K Seifert, and J M Moldowan, in *Advances in Organic Geochemistry* (eds A G Douglas, and J R Maxwell), Pergamon Press, Oxford, **1980**, pp 229-237
- 3 (a) P Bisseret, M Zundel, and M Rohmer, *Eur J Biochem*, **1985**, *150*, 29, (b) M Rohmer, and G Ourisson, *Tetrahedron Lett*, **1976**, 3641, (c) M Zundel, and M Rohmer, *Eur J Biochem*, **1985**, *150*, 35, and references cited therein
- 4 (a) R E Summons, and L L Jahnke, *Geochim Cosmochim Acta*, **1990**, *54*, 247, and references cited therein, (b) R E Summons, and L L Jahnke, in *Biological Markers in Sediments and Petroleum A tribute to Wolfgang Seifert*, Prentice Hall, in the press, (c) J Rullkötter, R Littke, and R G Schaefer, in *ACS Symposium Series N°429 Geochemistry of Sulphur in Fossil Fuels* (eds W L Orr, and C M White), copyright by the American Chemical Society, **1990**, pp 149-169
- 5 J Connan, *Bull Centres Rech Explor-Prod Elf-Aquitaine*, **1988**, *12*, 759
- 6 P Bisseret, and M Rohmer, *Tetrahedron Lett*, **1990**, *31*, 7445
- 7 R E Corbett, and C K Heng, *J Chem Soc C*, **1971**, 1885
- 8 R E Corbett, and R A J Smith, *J Chem Soc C*, **1967**, 1622
- 9 For a recent utilization of SOCl₂ as a dehydrating agent of tertiary alcohols, see I Farkas, and H Pfander, *Helv Chim Acta*, **1990**, *73*, 1980
- 10 H Ageta, K Shiojima, and Y Arai, *Chem Pharm Bull*, **1987**, *35*, 2705
- 11 Y Tsuda, K Isobe, S Fukushima, H Ageta, and K Iwata, *Tetrahedron Lett*, **1967**, 23
- 12 W J Dunstan, H Fazakerley, T G Halsall, and E R H Jones, *Croat Chem Acta*, **1957**, *29*, 173
- 13 I Kuwajima, and E Nakamura, *J Am Chem Soc*, **1975**, *97*, 3257
- 14 H Budzikiewicz, J M Wilson, and C Djerassi, *J Am Chem Soc*, **1963**, *85*, 3688
- 15 J-M Lehn, and G Ourisson, *Bull Soc Chim*, **1963**, 1113
- 16 G D Abbott, G Y Wang, T I Eglinton, A K Home, and G S Petch, *Geochim Cosmochim Acta*, **1990**, *54*, 2451
- 17 M Rohmer, C Anding, and G Ourisson, *Eur J Biochem*, **1980**, *112*, 541